Supplementary Information

Large-scale *in-vitro* production, refolding and dimerization of PsbS in different microenvironments

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Figure S1: Inclusion body purification from 200 mL cell pellet.

(+) IPTG sample shows the overexpression of PsbS from 200 mL culture. Several washes of inclusion body purification were carried out (supernatant S1 to pellet P4). Bovine serum albumin protein $(1.5 \, \mu g)$ is shown for comparison of yield. In the final step, pellet P4 contains PsbS of ~19mg, which was used for the next step, urea wash purification.

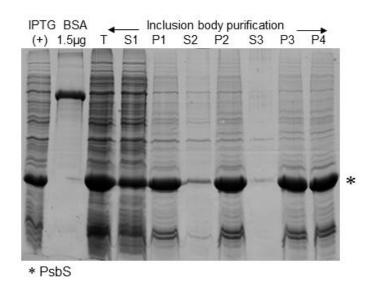


Figure S2 Purification of Lhcb1 (*Arabidopsis thaliana*) from inclusion body pellet using the urea wash protocol.

Lane 1: Lhcb1 from the inclusion bodies pellet was dissolved in buffer containing 8M urea. Several washes of urea buffer were carried out (Lane 2,3,4). Lane 5 is washing step of pellet with 8M urea buffer with 0.05% Lithium dodecyl sulfate (LDS). The last wash step was carried out using urea buffer with 0.5% of LDS to dissolve all the PsbS from inclusion bodies (Lane 6). Lane 7 contains 3µg of bovine serum albumin for yield comparison.

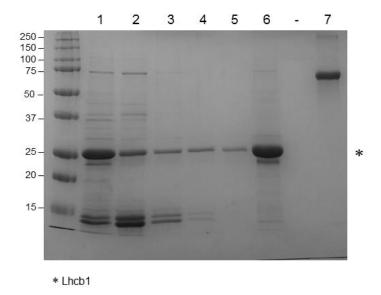


Figure S3 Homology model of dimeric PsbS from Physcomitrella patens

A homology model of *Physcomitrella patens* PsbS constructed by SWISS-MODEL based on the crystal structure of PsbS from *Spinacia oleracea* (PDB-ID 4RI2). The homology structure includes the stromal loops that are not resolved in the crystal structure and was energy-minimized using Chimera software¹. (a) front view of *Physcomitrella patens* PsbS (b) side view of *Physcomitrella patens* PsbS.

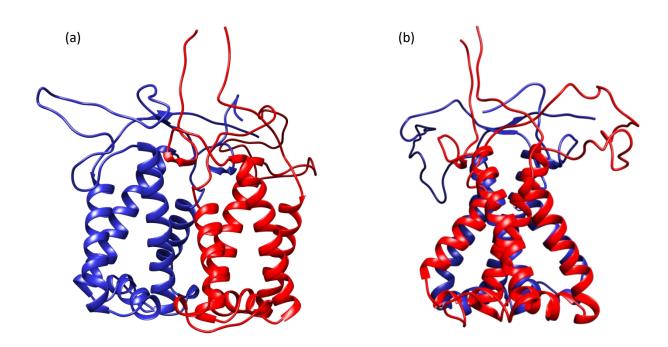


Figure S4: Boiled (B) and unboiled (UnB) samples of PsbS in FC 12.

Using the standard (sodium dodecyl sulfate) SDS-page gel protocol including sample boiling, both monomer and dimer bands of PsbS are observed in a 90-days old sample. If the boiling step before loading is omitted, only dimer bands are observed.



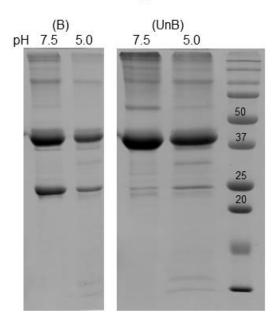


Figure S5: Comparison of size exclusion chromatograms of PsbS in n-Octyl- β -D-Glucopyranoside (OG) at pH 7.5 with the detection wavelength set at 260 nm (solid), and at 214 nm (dash) detection.

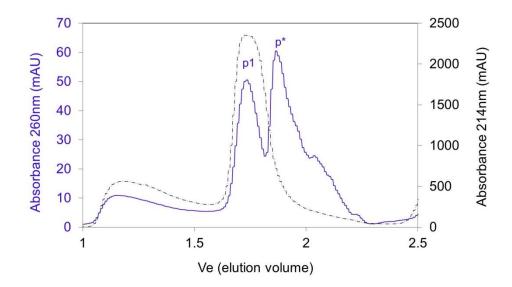


Table S1: Molecular weight estimates for SEC eluted fractions of PsbS in n-Dodecyl phosphocholine (FC-12)

Sample Description	Ve	MW (kDa)	Ехр	Y
PsbS pH 7.5 peak *	1.92	50	0.00135	50.6531
PsbS pH 7.5 peak 1	1.75	90	0.00243	90.9036
FC-12 detergent	1.98	41	0.001101	41.2066
PsbS pH 5.0 peak *	1.95	45	0.00122	45.6864
PsbS pH 5.0 peak 1	1.65	128	0.003427	128.226

Table S2 Molecular weight estimations for SEC eluted fractions of PsbS in OG

Sample description	Ve	MW (kDa)	Ехр	Y
PsbS pH 7.5 peak *	1.87	60	0.00186	60.6579
PsbS pH 7.5 peak 1	1.72	100	0.003081	100.439
OG detergent	1.92	51	0.001573	51.2722
PsbS pH 5.0 peak *	1.89	56	0.00174	56.7134
PsbS pH 5.0 peak 1	1.78	82	0.002518	82.0911

Figure S6 Calibration curve in FC12 (a) and in OG (b).

Markers 1, 2, 3 and 4 correspond to α -amylase (200KDa), alcohol dehydrogenase (150KDa), bovine serum albumin (66KDa) and carbonic anhydrase (29KDa) (green diamond).

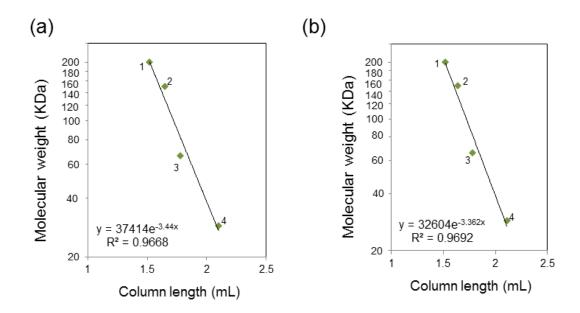


Fig S7: Western blot of refolded PsbS using Anti-PsbS (Agrisera antibodies AS09533)

- (a) SDS-page gel containing PsbS refolded in FC-12.
- (b) Refolded PsbS was run on western blot using Anti-PsbS antibodies to detect the presence of PsbS protein. The presence of dimer, monomer and higher aggregates are observed.

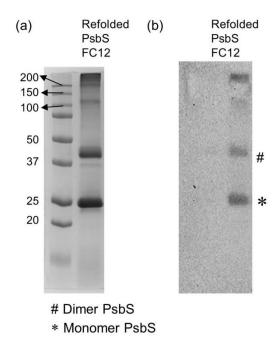
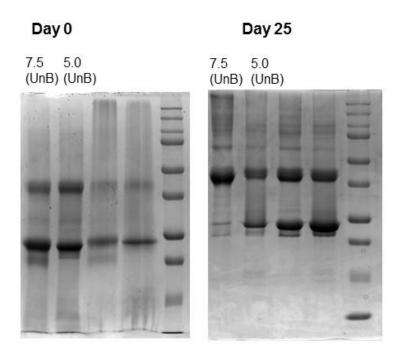


Fig S8: Original SDS-page gel figure graphs that were presented in edited form (cutting two lanes) in Figure 4 in the main text.



References

1. Pettersen, E. F. *et al.* UCSF Chimera — A Visualization System for Exploratory Research and Analysis. (2004). doi:10.1002/jcc.20084